

THE ROLE OF SIALIC ACID IN NATURAL MUD IN THE GROWTH AND NEURAMINIDASE SECRETION OF *VIBRIO CHOLERAE non-O1* STRAINS

I. Abrashev, P. Petrov, R. Eneva

Abstract: The growth rate and multiplication of *Vibrio cholerae non-O1* strain in the mud from the Pomorie lake (near the Black sea) were studied. The neuraminidase enzyme secretion and its accumulation at different growth phases and different cultivation temperatures were studied too. The obtained results demonstrate that the mud from the Pomorie lake is a good vibrio growth medium and substrate for the neuraminidase enzyme secretion. The role of the neuraminidase enzyme in the metabolism of mucins and glycoproteins on trophic level was confirmed.

Keywords: *Vibrio cholerae non-O1*, neuraminidase, mud.

1. INTRODUCTION

The representatives of genus *Vibrio* are natural inhabitants of the aquatic environment. Some species are pathogenic for humans. *V. cholerae* strains belonging to serotypes O1 and O139 cause cholera, a severe epidemic disease, resulting in dehydration and death (Kaper et al., 1995). *V. cholerae* strains, possessing the genetic potential to cause epidemic diseases are generally referred to as non-O1/non O139 *V. cholerae*.

In recent years, sporadic cases or limited outbreaks caused by non-O1/non O139 *V. cholerae* (Bhattacharya et al., 1998; Morris, 1990) have been reported. *V. cholerae* is part of the natural bacterial flora of the aquatic medium, including fresh, brackish and salty waters (Colwell and Hug, 1994). The ecological changes in the sea and estuarine medium provide good conditions for dissemination of *V. cholerae* and further the appearance of new epidemic strains, spread by birds and wastewater (Ruiz et al., 2000). The elucidation of mechanisms of pathogenic microorganisms' adaptation to the changes of environment conditions is a question of practical and theoretical interest. This depends on the genetic and biochemical abilities of bacteria with pathogenic potential to inhabit objects of the environment at various temperatures and limit of the available nutrition substances (Reid et al., 2002). Neuraminidase is an enzyme found in a number of viruses and bacteria, including *V. cholerae* (Burnet and Stone, 1947) and *V. cholerae non-O1* (Muller and Lutticken, 1974).

Bacterial neuraminidases are induced by a number of low- and high-molecular weight substances all of which possess terminal non-reducing N-acetyl-neuraminic groups. A number of natural substrates as soil, mud, water, etc., which contain mucins are suitable substrates for induction of the enzyme, as its secretion at the saprophytic free-living vibrios and aeromonads takes part in the metabolization of mucins and glycoproteins at trophic level.

The aim of the present work was to investigate the role of the Pomorie lake mud as a natural substrate containing mucins in the growth and neuraminidase production of *V. cholerae non-O1* in a wide temperature range and its relation to the growth phases.

2. MATERIAL AND METHODS

2.1. Bacterial strains

Research experiments were carried out with 17 *V. cholerae non-O1* strains of a different origin, pathogenic potential and neuraminidase production from the collection of the National Center of Infectious and Parasitic diseases in Sofia, kindly provided to us by associated professor, Doctor T. Kantardjiev. Tryptic soy broth (TSB) (Difco, Detroit, MI, USA), supplemented with 1% NaCl and pH 8.2, was used as a growth medium for the studied strains.

2.2. Cultivation conditions

The 24-hour culture was resuspended in phosphate buffer to bacterial suspension with density 1×10^6 cells/ml. Flasks containing 50 ml sterile Pomorie lake mud were inoculated with 1 ml bacterial suspension. Flasks containing 50 ml

nutrient broth supplemented with 0.1% glucomacropeptide and inoculated with 1 ml bacterial suspension were used as control cultures. The bacterial cultures were incubated at 4 °C, 25 °C and 37 °C for 72 hours. The cell quantity was determined by serial dilutions and counting of colonies on tryptic soy agar with pH 8.2 (Difco, Detroit, MI, USA). The results were presented

as the arithmetic mean of two experiments, each performed twice.

2.3. Mud from the Pomorie lake.

Mud from the Pomorie firth lake (near the Black sea), sterilized in autoclave at 121 °C for 15 minutes, was used as a growth medium and inducer for the neuraminidase enzyme secretion.

Table 1. Content of free and bounded sialic acids in mud from Pomorie lake.

Pomorie mud	Bounded sialic acids (µg/ml)	Free sialic acids (i g/ml)
Sterile mud	0.320 – 0.345	0.175 – 0.210
Non-sterile mud	0.290 – 0.320	0.160 – 0.175

2.4. Neuraminidase activity.

The neuraminidase activity was determined quantitatively according to Aminoff (1961). Glucomacropeptide (Abrashv et al., 1979) was used as a substrate and inducer. We assumed

one unit (U) of neuraminidase activity as the amount that releases 1 µg of sialic acid from the glucomacropeptide substrate for 1 minute under standard conditions.

Table 2. Neuraminidase activity of *V. cholerae* non-O1 strains isolated from different sources.

Strain	Source	Group according to Heiberg	Neuraminidase activity (U/ml)
V.cholerae non-O1 30	River water	I	20
V.cholerae non-O1 29	River water	II	28
V.cholerae non-O1 14	River water	II	26
V.cholerae non-O1 15	River water	II	24
V.cholerae non-O1 17	River water	II	19
V.cholerae non-O1 16	River water	II	21
V.cholerae non-O1 7	River water	I	17
V.cholerae non-O1 42	River water	I	23
V.cholerae non-O1 19	Sewage	I	25
V.cholerae non-O1 13	Sewage	II	22
V.cholerae non-O1 6	Sewage	II	18
V.cholerae non-O1 27	Sewage	II	27
V.cholerae non-O1 18	Sewage	I	23
V.cholerae non-O1 8	Seawater	I	24
V.cholerae non-O1 9	Seawater	IV	22
V.cholerae non-O1 11	Seawater	III	21
V.cholerae non-O1 1	Seawater	II	25

3. RESULTS AND DISCUSSION

A precondition of these investigations was the suggestion of Müller (1974) that a number of natural substrates as soil, mud and water containing mucins are a suitable substrate for induction of the neuraminidase enzyme. The enzyme secretion in saprophytes, free-living vibrios and aeromonads effects on the metabolization of mucins and glycoproteins at trophic level. We studied the effect of

different temperatures and the presence of inducer in the medium on the growth, multiplication and enzyme secretion of *V. cholerae* non-O1 in a model system containing firth mud from Pomorie lake. Previous to our studying the role of the firth mud it was tested for presence of sialic acids (Table 1). It appears from the table that the bounded sialic acids content varies from 0.320 µg /ml to 0.345 µg /ml in sterile mud and from 0.290 µg /ml to

0.320 μg /ml in non-sterile mud. Meanwhile, the content of free sialic acids was studied and it was considerably lower – from 0.175 μg /ml to 0.210 μg /ml in sterile mud and from 0.160 μg /ml to 0.175 μg /ml in non-sterile mud. There are no data in the literature available to us about presence of

sialic acids in lake sediments. There are no data about the role of the sialic acids in the behaviour of pathogenic microorganisms in natural substrates. The presence of sialic acids in the firth mud is essential, because they are inducer for the biosynthesis of neuraminidase enzyme.

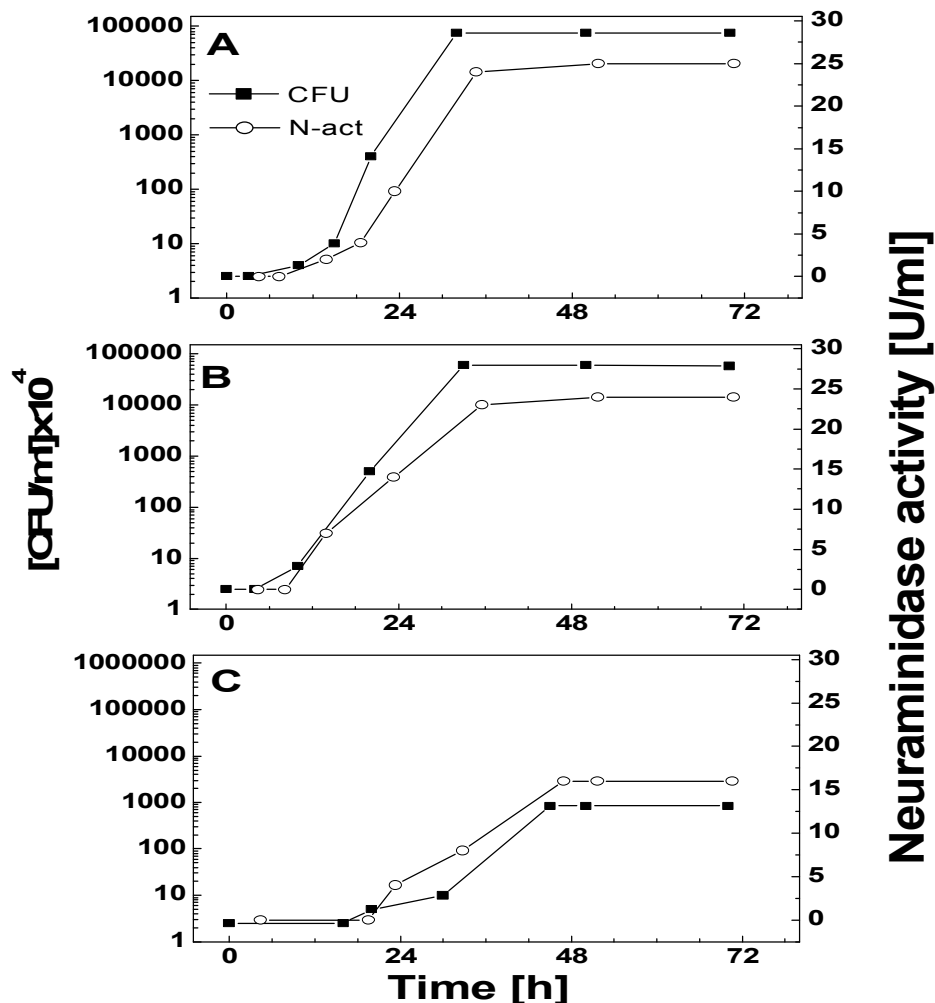


Fig. 1. Growth and neuraminidase activity of *V. cholerae* non-O1 29 strain cultivated in nutrient broth at 37°C (A), 25°C (B) and 4°C (C).

In view of choosing a proper enzyme producer, we carried out a screening of 17 *V. cholerae* non-O1 samples isolated from different sources of environment – river, sewage and seawater (Table 2). All the 17 samples produced neuraminidase activity between 17 and 28 U/ml. There was no statistically important difference in the neuraminidase activity levels between the isolates from different sources and the type according to Heiberg ($p > 0.05$). *V. cholerae* non-O1 29 strain appeared to be the best producer (28 U/ml) and this determined its following investigation. The re-

sults from the experiments on enzyme biosynthesis at each growth phase of this strain are represented on Fig. 1 – in broth and on Fig. 2- in firth mud. It was found that the lag-phase of the broth culture at 37 °C continues to the 3rd hour and no neuraminidase was produced during this period. The enzyme synthesis starts at the beginning of the log-phase, increases quickly, reaches its maximum (25 U/ml) at the 32nd hour and keeps this level till the 72nd hour of incubation. The lag-phase at 25 °C and 4 °C was 4 and 16 hours respectively, as enzyme synthesis in this period was not observed

too. The maximum enzyme secretion was observed in the end of the log-phase and in the be-

ginning of the stationary phase, 24 U/ml and 16 U/ml respectively.

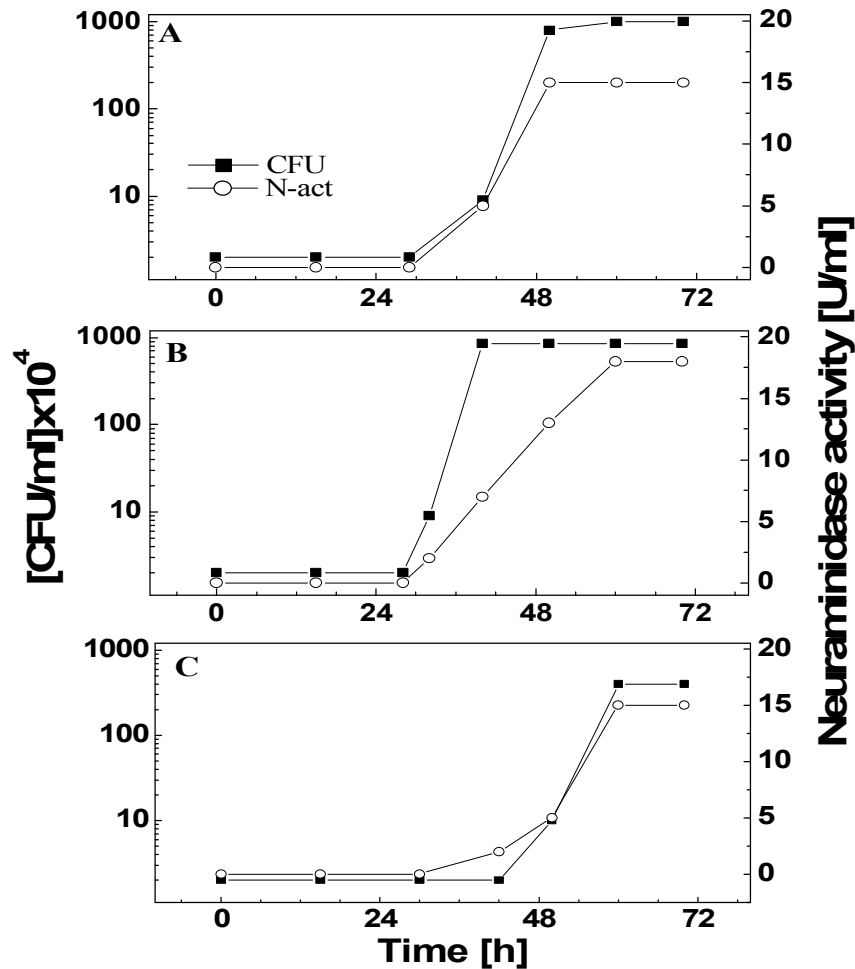


Fig. 2. Growth and neuraminidase activity of *V. cholerae* non-O1 29 strain cultivated in mud from Pomorie lake at 37°C (A), 25°C (B) and 4°C (C).

4. CONCLUSION

The results show that the Pomorie lake mud is an appropriate natural substrate for the vibrios growth, multiplication and neuraminidase secretion. Results obtained elucidate the metabolism of mucins and glycoproteins at trophic level and of

preserving some pathogenicity determinants as neuraminidase enzyme as well.

Acknowledgements: This work was supported by grants from Bulgarian Ministry of Sciences and Education, Science Research Fund G-2/2005.

REFERENCES

1. Abrashev I., Velcheva P., Nikolov P., and Kourteva J. (1979), Substrate for colorimetric determination of enzyme activity. Bulgarian patent Reg. 1 47647, AC 1 30580.
2. Abrashev I. and Orozova P. (2005), Influence of mucins in natural mud from Pomorie salty lake on the growth and neuraminidase secretion of some *Vibrio cholerae* non-O1 strains. *Compt. Rend. l'Acad. Bulg. Sci.* 58, 1213-1216.

3. Aminoff D. (1961), Methods for quantitative estimation of N-acetyl-neuraminic acids and their application to hydrolysates of sialomucoids. *Biochem. J.* 81, 384-392.
4. Bhattacharya M. K., Dutta D., Bhattacharya S. K., Deb A., Mukhopadhyay A. K., Nair G. B., Shimada T., Takeda Y., Chowdhury A., and Mahalanabis D. (1998), Association of disease approximating cholera caused by *Vibrio cholerae* of serogroups other than O1 and O139. *Epidemiol. Infect.* 120, 1-5.

5. Burnet F. M. and Stone J. D. (1947), The receptor destroying enzyme of *Vibrio cholerae*. *Austral. J. Exp. Biol. Med. Sci.* 25, 227-233.

6. Colwell R. R. and Hug A. (1994), *Vibrios in the environment: viable but not culturable Vibrio cholerae*. In: *Vibrio cholerae and cholera: Molecular to global perspectives* (eds. Wacshmith I. K., Blake P. A., and Olsvic O.), Washington, DC: American Society for Microbiology, 117-130.

7. Kaper J. B., Morris J. G. Jr., and Levine M. M. (1995), *Cholera*. *Clin. Microbiol. Rev.* 8, 316.

8. Morris J. G. Jr. (1990), Non-O group 1 *Vibrio cholerae*: a look at the epidemiology of an occasional pathogen, *Epidemiol. Rev.* 12, 179-191.

9. Müller H. (1974), Neuraminidases of bacteria and protozoa and their pathogenic role. *Behring Inst. Mitt.* 55, 34-56.

10. Müller H. and Lütticken R. (1974), Das Vorkommen von neuraminidase und N-acylneuraminat-Liase bei NAG-Vibrionen. *Path. Microbiol.* 41, 233-239.

11. Reid J. B., Stone P. J., Pearson A. J., and Wilson D. R. (2002), Yield response to nutrient sup-

ply across a wide range of conditions. 2. Analysis of maize yields. *Field Crops Res.* 77, 173-189.

12. Ruiz G. M., Rawlings T. K., Dobbs F. C., Drake L. A., Mullady T., Huq A., and Colwell R. R. (2000), Worldwide transfer of microorganisms by ships. *Nature* 408, 49-50.

13. Russell R. B., Sasieni P. D., and Sterner M. J. (1998), Supersites within superfolds: Binding site similarity in the absence of homology. *J. Mol. Biol.* 282, 903-918.

14. Uchiyama H., Todoroki T., and Matsui S. (1989), Effects of salinity on the survival of non-O1 *Vibrio cholerae* under environments of low temperature. *Kansenshogaku Zasshi* 63, 138-144. (In Japanese).

15. Uchiyama H. (1998), Survival of *Vibrio cholerae* non-O1 in river sediment during cold season. *Kansenshogaku Zasshi* 72, 218-222. (In Japanese).

16. Vertiev Y., Ezepechuk Y., Abrashev I., Khorlin A., and Krasnova I. (1975), Purification and characterization of neuraminidase produced by NAG-*Vibrio*. *Bioorg. Chem.* 1, 1639-1645. (In Russian).

Ignat Abrashev

Bulgarian Academy of Sciences, The Stephan Angeloff Institute of Microbiology, Department of Microbial Biochemistry, 26 Acad. G. Bonchev Str., 1113 Sofia, Bulgaria. Fax: ++ 359 28 7001 09. E-mail: abrashev@microbio.bas.bg

Petar Petrov

National Center of Infectious and Parasitic Diseases, Department Microbiology, 26 Yanko Sakazov Blvd, 1504 Sofia, Bulgaria.

Roumiana Eneva

Bulgarian Academy of Sciences, The Stephan Angeloff Institute of Microbiology, Department of Microbial Biochemistry, 26 Acad. G. Bonchev Str., 1113 Sofia, Bulgaria.

Ēāī àò Āáðàøāā

Áúéääðñèà Āèääāī èý íà Í áóèèòà, Ēí ñòèòò ìî ì èèðī àèī ēī āèý “Ñòáòáí Āí áāēī ā”, Ñāèòèý “Ī èèðī áí à áēī òèī èý”, òè. “Āèää. Ā. Āí í ÷āā” 26, 1113 Ñī òèý, Áúéääðñèý, òàēñ +359 28 7001 09, E-mail: abrashev@microbio.bas.bg

Ī àòúð Ī àòðī à

Í áóèī í àēāí òáí úúð ìî çàðàçí è è ì àðàçèðī è áí - èāñòè, ñāèòèý “Ī èèðī àèī ēī āèý”, 1504 Ñī òèý, áóè. Bí ēī Ñāèúçī ā 26

Đóī ýí à Āí áāà

Áúéääðñèà Āèääāī èý íà Í áóèèòà, Ēí ñòèòò ìî ì èèðī àèī ēī āèý “Ñòáòáí Āí áāēī ā”, ñāèòèý “Ī èèðī áí à áēī òèī èý”, òè. “Āèää. Ā. Āí í ÷āā” 26, 1113 Ñī òèý

ĐĪ ĒĒ Í À ÑĒĀĒĪ ĀĀÒĀ ĒĒÑĀĒĒÍ À Ā ĀÑÒĀÑÒĀĀÍ À ÒĒÍ Ò Ī ĐĒ ĐĀÑÒĀĒĒĀ Ē Í ĀÓĐĀĪ ĒÍ ĒĀĀÇÍ ĀÒĀ ÑĀĒĐĀÓĒĒ Í À ÛĀĪ Ī ĀĀ *VIBRIO CHOLERAЕ non-O1*

Ē. Āáðàøāā, Ī. Ī àòðī à, Đ. Āí áāà

Đàçpī ā: Ī ðī ó-áí è ñā ðāñòāæā è ðàçī í í æāāáí àòī í à Ûāī *Vibrio cholerae non-O1* ā èàè ì ò Ī Ī Ī ðèēñēī òī àçāðī (áèèçī āī ×āðī Ī Ī ðā). Ī ðī ñèāāáí à ā ñúĪ ñāèðāòèýòā í à áí çèī à í áòðāī èí èāāçā è í āāí āī òī í àò-ðóī āāí à Ī ðāç ðāçèè-í èòā ðāñòāæī è òāç è ðāçèè-í è òāī í áðāòòðè í à èóèðèèðāí ā. Ī í èó-áí èòā ðāçòèòāðè Ī Ī èāçāò, ÷ā èāèòā í à Ī Ī Ī ðèēñēī òī àçāðī ā āī áðā ñðāāā çā ðāçāèòèā í à àèāðèī í èòā. Ñèāēī àèòā èēñāèēī è, ñúāúðæāĪ è ñā í áý, ñā èí áóèóī ð çā ñāèðāòèýòā í à áí çèī à í áòðāī èí èāāçā. Ī í òāúðāāí à ā ðī èýòā í à òī çè áí çèī ā Ī àòāí èèçī à í à Ī òóèí èòā è àèèēī Ī ðī òāèí èòā í à òðī òè-í Ī í èāī.